

جمهورية العراق وزارة التعليم العالي والبحث العلمي جامعة ميسان كلية العلوم قسم الكيمياء

Poly (2-Hydroxyethyl Methacrylate) In medical application

بعث مقدم من قبل

الطالبة

الطالبة

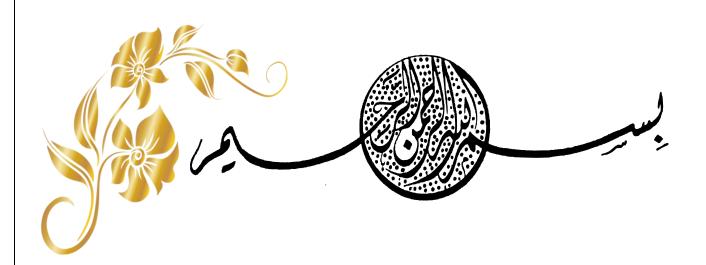
ناطمة بحبي حبر

زهراء راضي حسين

بحث مقدم الى مجلس كلية العلوم قسم الكيمياء / جامعة ميسان وهو جزء من متطلبات نيل شهادة البكالوريوس في قسم الكيمياء

<u>أشراف</u> م.د مثنی توفیق جوی

№ 2024 **№** 1445



إِنَّ فِي خَنْقِ السَّمَاوَاتِ وَالْأَرْضِ وَاخْتِلَافِ اللَّيْلِ وَالنَّهَارِ
وَالْقُلْكِ الَّتِي تَجْرِي فِي الْبَحْرِ بِمَا يَنْفَعُ النَّاسَ وَمَا أَنْزَلَ اللَّهُ
مِنَ السَّمَاءِ مِن مَّاءٍ فَأَحْيَا بِهِ الْأَرْضَ بَعْدَ مَوْتِهَا وَبَثَ فِيهَا مِن
كُلِّ دَابَّةٍ وَتَصْرِيفِ الرِّيَاحِ وَالسَّحَابِ الْمُسَخَّرِ بَيْنَ السَّمَاءِ
كُلِّ دَابَّةٍ وَتَصْرِيفِ الرِّيَاحِ وَالسَّحَابِ الْمُسَخَّرِ بَيْنَ السَّمَاءِ
وَالْأَرْضِ لَآيَاتٍ لِقَوْمٍ يَعْقِلُونَ

صدق الله العلي العظيم

سورة البقرة: 164





نهدي بجثنا المتواضع هذا....

الى سيد الكائنات . . . حبيبنا وشفيعنا . .

محمد (صل الله عليه وال وسلم)

الى امير المؤمنين . . . وقائد الغر المحجلين . . . علي (علية

السلام

الى من افتخر بجمل اسمة والدنا الحبيب . . . الى من صبرت على

الصعاب . . . نور عيني . . . والدتنا . . . الى . . . جميع من

ساندنا

الى من شاركتا في جهدنا المتواضع هذا . . . الدكتور الفاضل

(م.د مثنی توفیق جوي)

الشكر والعرفان

وبالشكر تدوم النعم

نشكر الله مولانا وخالقنا الذي من علينا بإتمام هذا العمل المتواضع مع رجائنا

ان يتقبله منا ويجعله خالصاً لوجه الكريم ..

وايماناً منا بفضل الاعتراف بالجميل فأننا نتقدم بخالص شكرنا وتقديرنا الى

((م.د مثنى توفيق جوي))الذي قبل مشكوراً الإشراف على هذا البحث،

وكان كريماً في توجيهه ونصحه فجزاه الله-تعالى - عنا وعن طلبة العلم خير

الجزاء . وكذلك نتقدم بالشكر الجزيل الى اساتذة قسم الكيمياء على ما

قدموه لنا من نصح ومشورة طيلة اعداد بحثنا المتواضع هذا ...

Abstract

Poly(2-hydroxyethyl methacrylate) (PHEMA) as a biomaterial with excellent biocompati ability and cytocompatibility elicits a minimal immunological response from host tissue making it desirable for different biomedical applications. This article seeks to provide an in-depth overview of the properties and biomedical applications of PHEMA for bone tissue regeneration, wound healing, cancer therapy (stimuli and non-stimuli responsive systems), and ophthalmic applications (contact lenses and ocular drug delivery.

As this polymer has been widely applied in ophthalmic applications, a specific consideration has been devoted to this field. Pure PHEMA does not possess antimicrobial properties and the site where the biomedical device is employed may be susceptible to microbial infections.

Therefore, antimicrobial strategies such as the use of silver nanoparticles, antibiotics, and antimicrobial agents can be utilized to protect against infections. Therefore antimicrobial strategies besides the drug delivery applications of PHEMA applications in the near future. The aim of this review was to bring together state of the art research on PHEMA and their applications.

1.Introduction

Poly(2-hydroxyethyl methacrylate) or PHEMA is a biocompatible, optically transparent, hydrophilic, and non degradable polymer. Morcover, it resists to crack propagation leading to a high load bearing ability [1-4] PHEMA is known to have a glass transmission (t) temperature and density in the range of 353-393 K. and 1.15-1.34 gml respectively [5], PIEMA is a biomaterial and a non-toxic polymer of the toxic monorer HEMA [6] PHEMA was initially studied and invented by Otto Wichterle and Drahoslav Lim in the development of modern soft hydrogel contact lenses in 1960 [2] PHEMA was first used as ar orbital implant, after which Wichterle and Lim thought the biological and mechanical properties of PHFMA made this biomaterial suitable for developing contact lenses [7]. Throughout the years, advancement in research and development has allowed PHEMA to be utilized in a broad range of biomedical applications. These the soft material contact lenses into four groups including norionic (low water content, <50%), nonionic (high water content, >50%), ionic (low water content, 50%), and ionic (high water contert, >50%) [12]. Table 1 represents the of **PHEMA** advantages and disadvantages applications

Table 1. Briefly summarizes the biomedical applications of PHEMA, advantages, and disadvantages.

Biomedical Applications	Advantages	Disadvantages	Reference
Bone tissue generation	Biocompatibility, excellent adhesion-promoting to other polymers, elastomeric properties	Non-degradable, requiring cross-linker	[10,13]
Wound healing	Transparency, biocompatibility	Low exudate absorbability, non- degradable, requiring cross-linker	[11,14]
Cancer therapy	Stimuli-responsive, inexpensive, easily combined with different polymers and drugs	Non-degradable requiring cross-linker	[15,16]
PHEMA hydrogel contact lens	Inexpensive, biocompatible, abundant copolymer possibilities	protein deposition issues	12

2-Application

2-1-Bone Tissue Regeneration

As bone is naturally composed of both organic and inorganic compounds, a combination of polymers and ceramics to mimic the bone's structure has always been a target.

The logic behind this combination is to bring the flexibility and rigidity of polymers and

ceramics into a package, respectively [31]. However, it is not an easy task, as essential requirements for a bone graft must be met, including biocompatibility, biodegradability, desirable mechanical properties based on the host tissue, osteoconductivity, osteoinductivity, and porosity [32]. Although PHEMA is endowed with great potential in biomedical engineering, it is a non-degradable polymer and, thus, many attempts have been made to address this issue. In an attempt to take the advantages of PHEMA in bone regeneration, it was used in combination with hydroxyapatite as a hybrid biocomposite hydrogel. monomer and cross-linker were evident to increase the degradation, while increasing in the bioceramic content decreased the weight loss; this phenomenon was attributed to the strong interaction between the ceramic particles and the anionic acrylic acid [38]. A ternary bone scaffold for bone regeneration was reported elsewhere, composed of polyurethane/PHEMA/cellulose nanowhiskers Notably, PHEMA was cross-linked with acrylic-urethane in situ in the presence of other phases. The increasing in the nanowhiskers was accompanied with a significant improve in the physicochemical and biological properties of scaffolds [13]. Besides using structural modifiers to make PHEMA degradable, blending this polymer with a degradable one would be another solution to its non-degradability Polycaprolactone (PCL) is known as an FDA approved biocompatible and degradable polymer with different applications in biomedical engineering. The main drawbacks of PCL, such as poor bioactivity and cell attachment, attributed to its hydrophobic nature [39]. Through a recent study, PHEMA was blended with PCL to obtain a degradable and hydrophilic hybrid. Moreover, to reinforce the mechanical properties of hybrid, a ceramic composed of hydroxyapatite and calcite was incorporated. Although no comparison was made between pure PCL and PCL/PHEMA, it is observable that the hybrid without the ceramic phase had about 20% water uptake and desirable cell attachment; moreover, the water contact angle was 89°. The obtained results implied that blending of both polymers culminated in the compensation of their restrictions—poor degradability and wettability [40]. As showcased, when it comes to designing a bone graft or scaffold, many requirements must be met. Tailorable physical and chemical properties of PHEMA would provide an opportunity to alter the final properties of bone graft based on the need and intended application.

2-2-wound Healing

The skin is formed mainly by three distinct layers, each of which has its own structure and function. The outer layer of skin is called the epidermis; the main duty of this layer is to protect the internal parts of the body from any potential danger from the outside, such as bacteria, contamination, etc. The protective role of the epidermis is accomplished through a 'brick and mortar'-like structure, in which corneccytes are bricks surrounded in non-polar lipids. The next layer is the dermis, mainly responsible for the mechanical support of skin tissue through the existence of extracellular matrix components. Moreover, different types of cells and compartments are located here, among which, fibroblasts outnumber the others as they can play a key role in the wound healing process and remodeling. The third layer is the hypodermis, which has important functions in the skin as the heat insulator and shock absorber [41]. Based on the depth, layer, and repair rate, wounds can be different from one another.

A full-thickness refers to a wound when all three layers are involved, and even deeper tissues, such as muscles and bone. When it comes to healing rate, the wounds are known as acute and chronic [42]. The former has a predictable time to heal completely and the time differs based on the wound severity, depth, etc. It complicates matters when the wound is diagnosed as chronic, because, in this case, the healing process gets stuck in one of the healing stages, hampering the process from completion. There are many reasons causing this process to prolong—immune-related diseases, diabetes,

infection, age, etc. These cases require more hospitalization and effective therapies [43]. Pivotal properties for ideal wound dressing include absorption of detrimental exudates, keeping the wound's moisture at an appropriate level, non-toxic nature, highly permeable to oxygen, anti-infection, and transparency. Because of high biocompatibility and low thrombogenicity, PHEMA has been widely used in ocular-related applications.

PHEMA hydrogels with a water content in the range of 32–43% are receiving considerable attention in wound healing applications. One recent study combined the great potential of PHEMA with bacterial cellulose whiskers, and silver, to design a multifunctional wound dressing. The cross-linking polymerization of HEMA was accomplished when the whiskerswere present. The whiskers significantly improved both water uptake and transparency simultaneously. The silver ions through the hydrogel showed acceptable antibacterial activity against both Gram-positive and Gram-negative bacteria. Besides the positive properties of prepared hydrogel, it was transparent, allowing to see the wound beneath the dressing and tracking healing on process. Based PHEMA, a thermosensitive polymer composed PHEMA/polynya/Cu was developed through free-radical polymerization for wound healing applications. The sol was "turned out" to undergo gelation when the temperature rose to 29 ° C. Designing such thermosensitive material is of particular interest because it can be injected through complex shape wounds, followed by being gelled. Therefore, shape processing and fabrication can be removed, making the wound dressing both cost-effective and efficient The addition of Cu to the polymer matrix culminated in both antibacterial activity and angiogenesis. The in vivo results implied that the ternary polymer decreased the healing time significantly [11]. Figure shows how the thermosensitive polymer was prepared and used as a wound dressing. Based on the water uptake of PHEMA, the wound dressing composed of this polymer benefited from absorbing the exudates, besides its great physicochemical and biological properties. Moreover, the transparency of PHEMA allows medical practitioners to easily track the wound healing process.

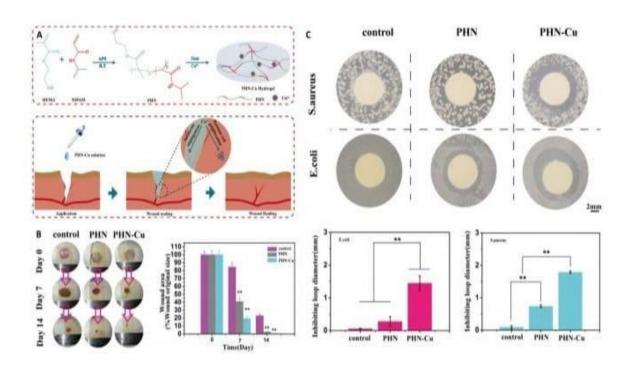


Figure 2. A schematic showing the preparation of thermosensitive polymer followed by wound healing (A); the in vivo results of different samples when they were applied on the wounds for 14 days (B); the antibacterial activity of samples against both Staphylococcus aureus and Escherichia coli bacteria (C); ammonium per sulfate (APS), radical,transformation (RT) poly-(HEMA-co-NIPAM) (PHN), reprinted from [11] with.permission from Wiley. ** denotes p < 0.01

2-3-Cancer therapy:

When referring to diseases with the highest causes of,mortality—cancer is second right after cardiovascular diseases [44]. The most common approaches to treat cancer are chemotherapy and radiation, which are capable of killing cancerous cells, but with catastrophic side effects for healthy cells. Newer methods:mainly focus on two points one involves decreasing the detrimental effects of chemotherapeutic drugs on healthy

cells and the other involves delivering the "cargo" to the targeted site. To achieve these aims, researchers have published a number of studies on these approaches in recent years Different drug delivery systems based on polymers, metals, ceramics, and composites were reported to not only present a but also be smart and responsive to disparate stimulants [45– 47]. An injection of drug molecules, with/without a carrier directly through the blood stream, is a common method in cancer therapy. When the drug molecules or drugloaded, nanoparticles enter the body they interact with endothelium in blood vessels to reach to the tumor site. If the nanocarrier is not responsive to any stimulant, its anticancer activity would fall into a passive targeting approach, which is mainly based on enhanced permeation and retention. Moreover, the release rate controlled release property, over an intended time period the type of material, structure, and physical and chemical properties, they can is based on the structure and design of the nanocarrier. Regarding release the drug through different mechanisms. On the other hand, active targeting uses some modifiers on the nanoparticle surface to "exactly" attach, this is followed by internalizing into the cancerous cell [48]. Moreover, there are other active targeting approaches based on internal and external stimulants. The internal-based stimuli responsive carriers are activated when they are exposed to the specific stimuli, such as pH, redox, ROS, etc. [49,50] the external

Responsive carriers can be controlled or activated through, different techniques. For instance magnetic nanoparticles respond to an external magnetic field to either reach an intended site and/or generate heat to eradicate the cancerous cells.through hyperthermia [51] Figure 3 exhibits the different drug delivery mechanisms towards cancer therapy

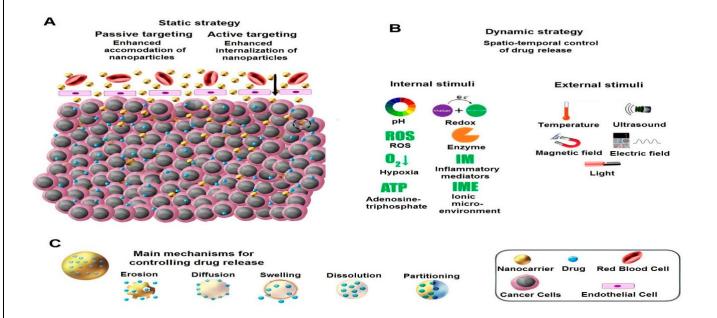


Figure 3. A-schematic showing different approaches toward drug delivery for cancer . ,therapy—static (A), dynamic (B)

and main mechanism for controlling drug release (C). Reprinted from [49] with permission from Elsevier but also be smart and responsive...

to disparate stimulants [45–47]. An injection of drug molecules, with/without a carrier directly through the blood stream, is a common method in cancer therapy. When the drug molecules or drug-loaded nanoparticles enter the body, they interact with endothelium in blood vessels to reach to the tumor site. If the nanocarrier is not responsive to any stimulant, its anticancer activity would fall into a passive targeting approach, which is mainly based on enhanced permeation and retention. Moreover, the release rate is based on the structure and design of the nanocarrier. Regarding the type of material, structure, and physical and chemical properties, they can release the drug through different mechanisms. On the other hand, active targeting uses some modifiers on the nanoparticle surface to "exactly" attach, this is followed by

internalizing into the cancerous cell [48]. Moreover, there are other active targeting approaches based on internal and external stimulants. The internal-based stimuli responsive carriers are activated when they are exposed to the specific stimuli, such as pH, redox, ROS, etc. [49,50]. The external responsive carriers can be controlled or activated through different techniques. For instance, magnetic nanoparticles respond to an external magnetic field to either reach an intended site and/or generate heat to eradicate the cancerous cells through hyperthermia [51]. Figure 3 exhibits the different drug delivery mechanisms towards cancer therapy.

exhibited a fast release at acidic medium, while the drug carrier was not supposed to be pH-responsive. The reason can be related to the cationic charge of doxorubicin as it altered the surface charge of nanoparticles resulting in an increase in the swelling properties at a lower pH [53]. A 5-Fluorouracil-loaded carrier composed of natural deep eutectic solvents and PHEMA was synthesized for cancer therapy. The physical structure was found porous with great potential for drug loading and sustained release. As expected, the encapsulation efficiency was approximately 77%, with a controlled release rate up to 130 h. The unloaded carrier showed no toxicity against HeLa cancer cells, whereas the anticancer-loaded sample effectively suppressed their proliferation [22]. The mentioned studies conducted research on hydrophilic drugs mostly, but via a report, curcumin as a hydrophobic drug was encap-sulated through PHEMA. The drug loading efficiency improved through gelled ionic liquid, obtaining a loading of 26.4%. The in vitro cell toxicity was assessed through ovarian cancer cells. The results implied that the capability of curcumin-loaded nanoparticles against the cancer cells was higher than pure curcumin [54]. To improve PHEMA functionality, it was combined with other polymers to yield a multifunctional drug delivery system for N', N- triacetic acid for positron emission tomography imaging. Doxorubicin was chosen as the anticancer drug and the release studies revealed that the drug liberation was pH-dependent. In vivo studies showed a higher accumulation of nanoparticles in tumors treated with 64Cu-labeled targeted micelles. Based on the results, the nanocarrier has a promising potential in theranostic-based cancer therapy

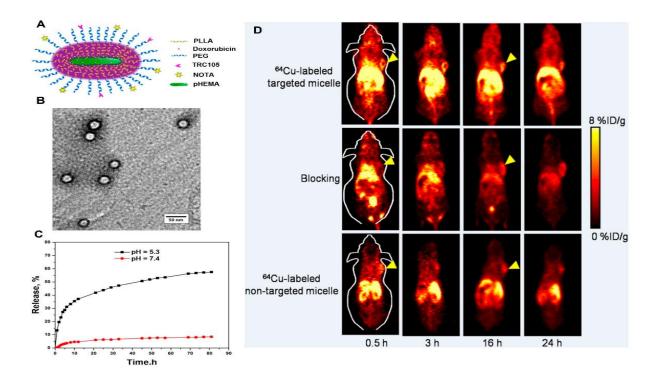


Figure 4. A schematic of the micelle (A); TEM micrograph of the micelle (B); the release rate of doxorubicin-loaded micelle at physiological and acidic mediums (C); the positron emission tomography images of 4T1 tumor-bearing mice treated with 64Cu-labeled targeted, non-targeted micelles, and targeted micelles with a blocking dose of TRC105 at different time intervals (D).

Reprinted from [55] with permission from American Chemical Society Nowadays, smart nanomaterials are at the forefront of every field of study as their efficiency and applicability are definitely evident compared to the conventional materials

As briefly discussed in the previous paragraph, smart nanoparticles in cancer therapy can be designed in a way to respond to a specific stimulus, e.g., temperature, pH, enzyme, redox, etc. In the case of cancer therapy, cancerous cells and tumor microenvironments have some specific features by which a smart drug delivery system can be triggered [47]. For instance, cancerous cells generate high amounts of hydrogen

sulfide, which can be used against cancerous cells to yield a smart delivery system; this stimulant's advantage was taken to synthesize a smart drug delivery system for cancer diagnosis and targeted therapy. A series of micelles composed of N-(2-hydroxyethyl)-4-azide-1,8-naphthalimide ended amphiphilic diblock copolymer poly(2-hydroxyethyl methacrylate)-block-poly(methyl methacrylate) (N3-Nap-pHEMA-b-PMMA-N3), loaded with doxorubicin, was designed and prepared. In the presence of hydrogen sulfide, the azido groups on the micelles reduced amid groups, culminating in green fluorescence and charge reversal. Moreover, the drug-loaded sample showed a faster release rate when exposed to hydrogen sulfide. Fluorescence imaging revealed that the drug-loaded micelles accumulated in the tumor's site selectively [16]. Figure 5 exhibits the applicability potential of hydrogen sulfide- responsive drug-loaded and non-loaded N3-Nap-pHEMA-b-PMMA-N3 micelles in cancer detection and therapy. It is wellknown that the tumor microenvironment has an acidic pH, and by having a pHresponsive delivery system that is protonated in an acidic medium, the cargo is liberated when the carrier reaches the tumor's microenvironment; thus, the hazardous effects of chemotherapeutic drugs on the healthy cells can be hampered [56,57]. Considering the swelling behavior and hydrophilic nature of PHEMA, it was used for improving the swelling behavior of N, N-dimethylaminoethyl methacrylate as a cationic polymer [58]. A pH-sensitive nanohydrogel delivery system was designed based on the combination of those polymers. At the physiological environment (pH = 7.4), no significant drug release was observed, whereas at acidic mediums, the faster release was evident. (cloud point changes from 28 °C to 39 °C (pH = 6.5)) [60]. An external-based stimuli-responsive nanocarrier composed of hydrophobic PMMA, hydrophilic PHEMA, and magnetite nanoparticles was designed for cancer therapy. Methotrexate was first grafted on the low molecular PHEMA followed by making a covalent bond between magnetite nanoparticles and the drug-grafted polymer. The main aim process (A), the TEM micrographs of materials obtained at each step of the synthesis process (B), the release rate (C), and the magnetic property of nanoparticles (D). The TEM micrograph of the Janus nanoparticle (Figure 6B(d)) clearly exhibits that there are two distinct areasvisible proving the successful synthesis of Janus structure. The obtained drug delivery system was turned out as a double-stimuli-responsive carrier—pH and magnetic—with great potential for cancer therapy.

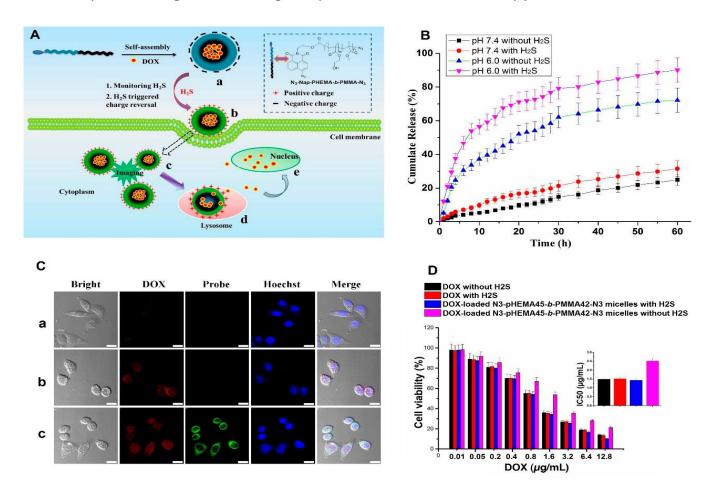


Figure 5. The mechanism of the drug-loaded hydrogen sulfide-responsive micelles when exposed to the stimulant is depicted schematically (A); the drug release rates of the drug-loaded N3-Nap-pHEMA-b-PMMA-N3 sample at acidic and physiological environments, with/without being exposed to hydrogen sulfide (B); the images of HeLa cells treated with doxorubicin (a), drug-loaded N3-Nap-pHEMA-b-PMMA-N3 without hydrogen sulfide (b), and drug-loaded N3-Nap-pHEMA-b-PMMA-N3 with hydrogen sulfide (c), taken by confocal microscopy (scale bar = $20~\mu m$) (C); the cell viability (%) of drug and non-loaded samples, with/without being exposed to hydrogen sulfide (D). Reprinted from [16], with permission from the American Chemical Society.

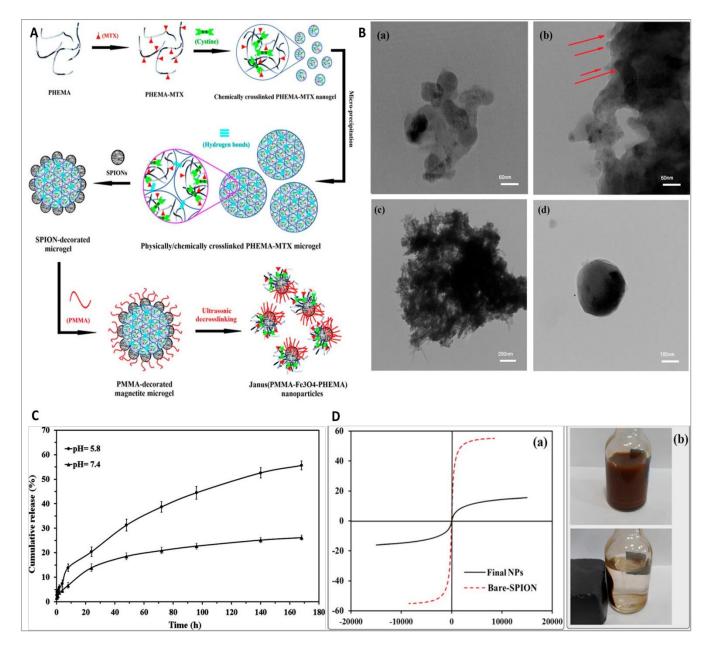


Figure 6. A schematic exhibiting the synthesis procedure of PHEMA-PMMA-magnetite (Fe3O4) (A); the TEM micrographs of the cross-linked PHEMA nanogel (chemically) (a), the cross-linked PHEMA microgel (physically and chemically) (b), Fe3O4- coated cross-linked PHEMA-methotrexate (c), pHEMA-PMMA-Fe3O4 (Janus) (d) (B); the release trends of methotrexate from PHEMA-PMMA-Fe3O4 nanoparticles (C); the magnetic hysteresis loops of bare Fe3O4 and final PHEMA-PMMA-Fe3O4 nanoparticles (a) plus separation of pHEMA-PMMA-Fe3O4 nanoparticles in water by a magnet (b) (D). Reprinted from [15], with permission from the American Chemical Society. (Red arrow: in Figure (b) show coagulation of cross-linked PHEMA nanogel).

3. Soft Contact Lenses

3. 1 Vision Correction

Contact lenses are ocular prosthetic devices widely used to correct vision. They can also be used for aesthetic or therapeutic purposes. A contact lens is a thin, curved plastic disk designed to be worn directly on the cornea for the correction of vision disorders of the eye. Like eyeglasses, contact lenses correct vision disorders caused by refractive errors whereby the eye is unable to refract light properly onto the retina, resulting in blurred images perceived by the brain [61,62]. Common types of refractive errors include astigmatism, myopia, hyperopia, and presbyopia. Besides blurred vision, refractive errors may also result in squinting, headaches, or eye strain. Contact lenses help to improve vision by correcting the light refraction path into the eye, thereby focusing light better on the retina. Unlike eyeglasses, contact lenses have the advantage of providing maximum peripheral

vision. This makes them more convenient and especially useful in sports. They are also useful in correcting vision when there is a huge difference in the power of each

PHEMA-based hydrogels are especially suitable in the fabrication of soft contact lenses. Besides being transparent, soft, and flexible, PHEMA-based hydrogels also exhibit superior biocompatibility, high oxygen permeability and sufficient mechanical strength [64]. Being soft and flexible aid in the comfort of wearers of soft contact lenses as opposed to rigid lenses, while the high oxygen permeability helps prevent adverse clinical events associated with corneal hypoxia. The mechanical strength of PHEMA-based hydrogels also ensures that soft contact lenses are reasonably durable

Three-dimensional (3D) printing of contact lenses for vision correction is currently in the research and development phase. In 2016, Johnson & Johnson Vision Care launched a collaboration with HP, Inc. to develop 3D printable contact lenses. Johnson & Johnson researchers are working with HP's 3D printing business team to utilize

smart and 3D print- ing technology to develop 3D printable contact lenses that can be personalized according to the needs of the patients and can be used to correct vision, reduce glare, eyestrain, and relieve eye allergies [68–70]. Therefore, 3D printing of high precision contact lenses for vision correction and ocular drug delivery is

3.2. Ocular Drug Delivery

Delivery of therapeutic drugs to the eyes through utilizing contact lenses fabricated from PHEMA-based hydrogel is a promising method to treat eye infections and disorders. Topical ocular drug administration can be used to treat superficial eye infections, such as conjunctivitis, blepharitis, and keratitis and intraocular disorders through corneal absorption such as uveitis and glaucoma [18,71].

Owing to the anatomical and physiological constraints of the eye, a major challenge of administering medication to the eye includes delivering an optimal concentration of the medicine to the active site for a necessary residence time so as to maximize the therapeutic effects. Drug delivery to the eye using drug-loaded hydrogel contact lenses enables a controlled and extended release of an optimal drug concentration by diffusion through the hydrogel system. This may improve ocular treatment efficacy by enabling higher bioavailability, prolonged residence time, lower systemic absorption, and improved patient compliance with dosage regimens [72]. Contact lenses loaded with drugs can provide treatment for ocular diseases and refractive errors at the same time, while patients without refractive error may utilize neutral lenses. Figure 7 displays the difference in drug release time using contact lenses and eye drops.

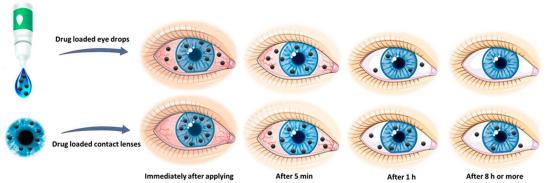


Figure 7. Schematic representation depicting the difference in drug release time using contact lenses as compared to the conventional topical formulation

Soft contact lenses fabricated from PHEMA-based hydrogels are especially suitable for ocular drug delivery. PHEMA-based hydrogels are soft, flexible, and highly oxygen permeable, and provide safety and comfort to patients during ophthalmic treatments. When a drug-loaded PHEMA-based hydrogel lens is applied onto the cornea of the eye, a thin film of tear is created, which gets trapped between the hydrogel lens and the cornea, resulting in the release of drugs through diffusion. The turnover duration of the tear film is longer compared to a normal tear film, resulting in a prolonged precorneal residence time. This results in higher bioavailability and a more accurate dose of the drug being delivered to the active site [61,73]. With the use of PHEMA-based lenses, patient compliance to dosage regimen can be more assured. In addition, the cost of ocular drug delivery by means of contact lenses is lower than repeated topical dosing with eye drops, hence reducing the cost of therapy. In 2007, Li and Chauhan conducted in vitro experiments to deliver timolol maleate through the use of PHEMA-based hydrogel contact lenses [74]. The experimental results showed that at least 20% of the drug that was entrapped in the lens will enter the cornea. This bioavailability is much greater than that of conventional topical dosing through eye drops.

3.3 Antimicrobial Strategies

Soft contact lenses synthesized from PHEMA-based hydrogels are widely used to correct refractive errors of the eyes and for sustained drug delivery to treat ocular diseases. Despite the benefits brought about by contact lenses, they are also the causes of eye infections and inflammatory episodes due to microbial contamination. These microbially driven adverse events range from microbial keratitis (MK), infiltrative keratitis (IK), contact lens-induced peripheral ulcer (CLPU), and contact lens-induced acute red eye (CLARE) [66]. Severe ocular infections and inflammation resulting from microbial contamination of contact lenses may lead to corneal injury resulting in vision impairment. Contamination of contact lenses can be attributed to various microbes, such as bacte- ria, parasites, viruses, and fungi. Gram-positive bacteria, such as streptococcus pneumoniae and Staphylococcus aureus, are predominantly responsible for the prevalence of contact lens-related CLPU and IK, while Gram-negative bacteria, such as pseudomonas aeruginosa, Haemophilus influenzae, and chlamydia trachomatis are predominantly responsible for the prevalence of CLARE [67]. Since pure PHEMA does not possess antimicrobial proper- ties against a broad range of gram-positive and gram-negative bacteria, antimicrobial approaches can be utilized to fabricate antimicrobial contact lenses to combat different microbes. These antimicrobial agents suppress or stop the growth of microbes through four modes of action: (1) penetrating directly into microbial cells, (2) modifying the microbial- substrate interfaces, (3) interfering with the quorum-sensing mechanism of microbial cells, and (4) generating reactive oxygen species [67]. Figure 8 illustrates different antimicrobial strategies for the development of antimicrobial contact lenses employing antibiotics, anti-fungal drugs, antimicrobial peptides, passive surface modifications, free-radical fabricating agents, quorum sensing quenchers, metals and metal oxide nanostructured materials, and coatings with antimicrobial nanomaterials.

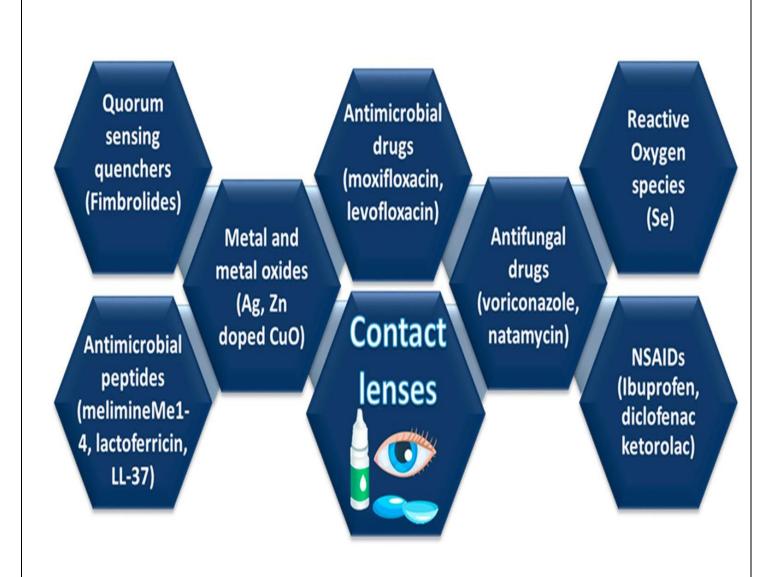


Figure 8. Antimicrobial strategies to fabricate microbiocidal contact lenses

Biomedical devices are widely used for the treatment of diseases and to replace injured or diseased organs or tissues in the human body. Since some of these devices are exposed to hostile external environments, they can potentially become contaminated with harmful pathogens and other infectious agents. This increases the possibility of developing an infection, which may lead to further complications, at the site where the biomedical device is employed. Therefore, adopting effective antimicrobial strategies is essential for the safe and effective use of biomedical devices to advance healthcare

[75]. Table 2 lists the different approaches for incorporation of antimicrobial drugs into PHEMA contact lenses.

Table 2. Various approaches and drugs used for contact lens drug delivery.

No.	Method of Drug Incorporation	Polymers/Contact Lens	Drugs Used	Inference	Ref
1	Soaking and lyophilization	pHEMA hydrogels containing cross-linked β-cyclodextrin-hyaluronan	Diclofenac	Improved the oxygen permeability, water uptake ability, surface hydrophilicity, and flexibility	[62]
2	Soaking method	HEMA, siloxane, ethylene glycol dimethacrylate, Irgacure [®] , dimethyl acrylamide	Timolol	Reduce burst release and the timolol retention time improved, the control release kinetics and uptake of timolol improved without varying the transmittance and swelling of the contact lens.	[76]
3	Polymerization of HEMA	HEMA, mPEG-PLA micelles	Latanoprost and Timolol	Improvement in drug residence time and higher bioavailability	[77
4	Free radical polymerization of the monomer by photoinitiation	pHEMA, chitosan nanoparticles	Dexamethasone Sodium Phosphate	Significantly enhanced bioavailability of ophthalmic drugs	[78
5	Free radical polymerization technique	pHEMA cross linked dextrin	Ciprofloxacin hydrochloride	Excellent physical stability as carrier for ciprofloxacin up to 3 months	[23
6	Bioinspired imprinted pHEMA- hydrogels	HEMA, zinc methacrylate, 1- or 4-vinylimidazole, and N-hydroxyethyl acrylamide	Acetazolamide or Ethoxzolamide	Remarkable improvement in the performance as controlled release system	[61
7	Prepared by photopolymerization	pHEMA, HEMA, mono-methacrylated β-CD (mono-MA-β-CD) and trimethylolpropane trimethacrylate	Puerarin	The data demonstrate that pHEMA/β-CD hydrogel contact lenses can effectively deliver puerarin through the rabbit's cornea	[79
8	β-CD was grafted to the gel network and soaked in drug solution	pHEMA and GMA	Diclofenac sodium	The hydrogels with pendant β-CD are particularly useful for the development of cytocompatible drug loaded SCLs	[80
9	By ultraviolet light polymerization film coating process	PLGA films over pHEMA	Ciprofloxacin, fluorescein	PLGA film coated over pHEMA lenses sustained the release of drug, which can be controlled by changing either the ratio of drug to PLGA or the molecular mass of the PLGA used	[71
10	Gels prepared by living /controlled imprinting technique	HEMA, acrylic acid, acrylamide and methacrylic acid	Ketotifen, diclofenac sodium	Imprinting via living polymerization extends or delays the template release profile by two-fold over that of imprinting via conventional free-radical polymerization techniques	[81

Table 2. Cont.

No.	Method of Drug Incorporation	Polymers/Contact Lens	Drugs Used	Inference	Ref.
11	Dispersion of microemulsion in contact lenses	рНЕМА	Hexadecane	p-HEMA gels loaded with a microemulsion, stabilized with silica shell are transparent and releases drugs for a period of over 8 days in vitro	[82]
12	Molecular imprinting technique	HEMA+ methacrylic acid or meth-methacrylate	Timolol	Incorporation of meth-methacrylate as comonomer increases the timolol loading capacity to therapeutically useful levels while retaining appropriate release characteristics in vitro	[83]
13	Molecular imprinting technique	HEMA + methacrylamide propyl trimethylammonium chloride	Azulene	Molecular imprinting is capable to store the anionic drug such as azulene based on ion-exchange reaction	[84]

Silver is recognized for its remarkable microbicidal properties against a broad spectrum of gram-positive, gram-negative, and multidrug-resistant bacteria, protozoa, and fungi [85]. Silver-infused polymers have been employed to fabricate commercially avail- able antimicrobial contact lens cases. Upon the addition of sterile contact lens solution to the cases, silver ions will be released into the solution at a sustained rate to kill the harmful microorganisms and to inhibit their growth [67]. The multiple biocidal actions of silver ions may also prevent contemporaneous bacterial strain resistance

[66]. The biocidal silver containing-agents are due to (i) they generate reactive oxygen species; (ii) the interfering with membrane protein, enzymes, and DNA; (iii) their interaction with the bacterial cell wall [85–87]. Antibiotics, such as tetracycline, erythromycin, chloramphenicol, and ciprofloxacin can also be used to fabricate antimicrobial contact lenses. Antibiotics work by directly destroying or inhibiting the growth of bacteria on the contact lens and the cornea. Qin et al. loaded contact lenses with ciprofloxacin to treat Pseudomonas aeruginosa bacterial keratitis in an ex vivo infection model, with the study showing good efficacy and with human

Other strategies to synthesize antimicrobial contact lenses include utilizing antimicrobial agents fabricated through other methods. These methods include generating reactive oxygen species, use of nonsteroidal anti-inflammatory drugs, fimbrolides (quorum-sensing quenchers), antimicrobial peptides, and passive non-cidal molecules [67,73,89,90]. While many different approaches to synthesize antimicrobial contact lenses and lens cases have been investigated, the most promising one involves using silver-infused contact lens and lens cases which are already available commercially. The other antimicrobial strategies, through continuous research, may also find commercial applicability in the near future.

4. Conclusion

The purpose of this review was to bring together state-of-the-art research on PHEMA and their applications. PHEMA is a biomaterial that has been utilized in many biomedical applications. PHEMA is rarely used in isolation and is usually utilized as a copolymer with other biomaterials in these applications. Initially employed in soft contact lenses due to its superior biocompatibility, unique physical and mechanical properties, the biomedical applications of PHEMA have since been extended to include hydrogel wound dressings, bone tissue regeneration, cancer therapy, ocular drug delivery, artificial skin, artificial cornea, breast augmentation, catheters, intrauterine inserts and prosthesis. However, since pure PHEMA is non-degradable, it has to be combined with other materials or modified based on the desire applications. Moreover, PHEMA does not possess inherent ant micro bail properties and so additives such as silver nanoparticles, antibiotics, and nonsteroidal anti-inflammatory drugs have to be used to give biomedical devices containing PHEMA their antimicrobial characteristics to protect against infections. The excellent biocompatibility, physicochemical, and cell-friendly properties of PHEMA also make it an attractive biomaterial in tissue engineering .

References

- .1 Roberts, J.J.; Martens, P.J. Engineering biosynthetic cell encapsulation systems. In Biosynthetic Polymers for Medical Applications; Elsevier Inc.: Amsterdam, The Netherlands, 2016; pp. 205–239, ISBN 9781782421139.
- .2 Barui, A. Synthetic polymeric gel. In Polymeric Gels; Elsevier: Amsterdam, The Netherlands, 2018; pp. 55–90.
- .3 Madigan, N.N.; Windebank, A.J. Spinal cord injury. In Principles of Tissue Engineering; Elsevier: Amsterdam, The Netherlands, 2020; pp. 1047–1091.
- .4 Fink, J.K. Grafting. In Reactive Polymers Fundamentals and Applications; Elsevier: Amsterdam, The Netherlands, 2013; pp. 425–452.
- .5 Poly (2-Hydroxyethyl Methacrylate). Available online: http://polymerdatabase.com/polymers/Poly2-hydroxyethyl-2- methylpropenoate.html (accessed on 23 May 2021.(
- .6 Kulygin, O.; Silverstein, M.S. Porous poly (2-hydroxyethyl methacrylate) hydrogels synthesized within high internal phase emulsions. Soft Matter 2007, 3, 1525–1529. [CrossRef] [PubMed]
- .7 Hicks, C.R.; Moris, I.T.; Vijayasekaran, S.; Fallon, M.J.; McAllister, J.; Clayton, A.B.; Chirila, T.V.; Crawford, G.J.; Constable, I.J. Correlation of histological findings with gadolinium enhanced MRI scans during healing of a PHEMA orbital implant in rabbits. Br. J. Ophthalmol. 1999, 83, 616–621. [CrossRef]
- Shery, J.; Anroop, B.N.; Jigar, S.; Nagaraja, S.; Sumeet, G.; Pottathil, S. Emerging Role of Hydrogels in Drug Delivery Systems, Tissue Engineering and Wound Management. Pharmaceutics 2021, 13, 357.
- .9 Caló, E.; Khutoryanskiy, V.V. Biomedical applications of hydrogels: A review of patents and commercial products. Eur. Polym. J.
-] .267–252 **.**65 **.**2015CrossRef[
- .10 Chen, L.; Tang, Y.; Zhao, K.; Liu, J.; Jiao, M.; Bai, H.; Wu, Z. HEMA-Modified Expandable P (MMA-AA) Bone Cement with Dual Water Absorption Networks. Macromol. Mater. Eng. 2020, 305, 1900752. [CrossRef[
- .11 Wang, Y.; Chu, X.; Sun, Y.; Teng, P.; Xia, T.; Chen, Y. A convenient approach by using poly-(HEMA-co-NIPAM)/Cu2+ solution sol-gel transition for wound protection and healing. J. Biomed. Mater. Res. Part B Appl. Biomater. 2021, 109, 50–59. [CrossRef[
- Musgrave, C.S.A.; Fang, F. Contact lens materials: A materials science perspective. Materials 2019, 12, 261. [CrossRef]

- .13 Shahrousvand, M.; Ghollasi, M.; Zarchi, A.A.K.; Salimi, A. Osteogenic differentiation of hMSCs on semi-interpenetrating polymer networks of polyurethane/poly (2-hydroxyethyl methacrylate)/cellulose nanowhisker scaffolds. Int. J. Biol. Macromol. 2019, 138, 262–271. [CrossRef]
- .14 Di, Z.; Shi, Z.; Ullah, M.W.; Li, S.; Yang, G. A transparent wound dressing based on bacterial cellulose whisker and poly (2-hydroxyethyl methacrylate). Int. J. Biol. Macromol. 2017, 105, 638–644. [CrossRef]
- .15 Ghanbarinia Firozjah, R.; Sadeghi, A.; Khoee, S. Ultrasonic De-cross-linking of the pH- and Magneto-Responsive PHEMA/PMMA Microgel to Janus Nanoparticles: A New Synthesis Based on "Grafting from"/"Grafting to" Polymerization. ACS Omega 2020, 5, 27119–27132. [CrossRef[
- Zhang, H.; Kong, X.; Tang, Y.; Lin, W. Hydrogen Sulfide Triggered Charge-Reversal Micelles for Cancer-Targeted Drug Delivery and Imaging. ACS Appl. Mater. Interfaces 2016, 8, 16227–16239. [CrossRef]
- .17 Bashir, S.; Hina, M.; Iqbal, J.; Rajpar, A.H.; Mujtaba, M.A.; Alghamdi, N.A.; Wageh, S.; Ramesh, K.; Ramesh, S. Fundamental concepts of hydrogels: Synthesis, properties, and their applications. Polymers 2020, 12, 2702. [CrossRef]
- .18 Chatterjee, S.; Upadhyay, P.; Mishra, M.; Akshara, M.R.; Sifat Zaidi, Z.; Iqbal, S.F.; Misra, S.K. Advances in chemistry and composition of soft materials for drug releasing contact lenses. RSC Adv. 2020, 10, 36751–36777. [CrossRef[
- .19 Halpenny, G.M.; Steinhardt, R.C.; Okialda, K.A.; Mascharak, P.K. Characterization of pHEMA-based hydrogels that exhibit light-induced bactericidal effect via release of NO. J. Mater. Sci. Mater. Med. 2009, 20, 2353. [CrossRef]
- .20 Macková, H.; Plichta, Z.; Hlídková, H.; Sedlác´ek, O.; Konefal, R.; Sadakbayeva, Z.; Dušková-Smrc´ková, M.; Horák, D.; Kubinová, Š. Reductively Degradable Poly (2-hydroxyethyl methacrylate) Hydrogels with Oriented Porosity for Tissue Engineering Applications. ACS Appl. Mater. Interfaces 2017, 9, 10544–10553. [CrossRef] [PubMed[
- .21 Gunatillake, P.A.; Adhikari, R. 2—Nondegradable synthetic polymers for medical devices and implants. In Woodhead Publishing

Series in Biomaterials; Poole-Warren, L., Martens, P., Green, R., Eds.; Woodhead Publishing: Cambridge, UK, 2016; pp. 33–62, ISBN 978-1-78242-105-4.

- .22 Pradeepkumar, P.; Subbiah, A.; Rajan, M. Synthesis of bio-degradable poly (2-hydroxyethyl methacrylate) using natural deep eutectic solvents for sustainable cancer drug delivery. SN Appl. Sci. 2019, 1, 568. [CrossRef]
- .23 Das, D.; Pal, S. Dextrin/poly (HEMA): PH responsive porous hydrogel for controlled release of ciprofloxacin. Int. J. Biol. Macromol.
-] .178–171 **.**72 **.**2015CrossRef[
- .24 Fuest, M.; Yam, G.H.; Mehta, J.S.; Duarte Campos, D.F. Prospects and Challenges of Translational Corneal Bioprinting.

Bioengineering 2020, 7, 71. [CrossRef] [PubMed]

- .25 Wang, J.; Karami, P.; Ataman, N.C.; Pioletti, D.P.; Steele, T.W.J.; Klok, H.-A. Light-Activated, Bioadhesive, Poly (2-hydroxyethyl methacrylate) Brush Coatings. Biomacromolecules 2020, 21, 240–249. [CrossRef] [PubMed[
- .26 Wang, C.G.; Chong, A.M.L.; Pan, H.M.; Sarkar, J.; Tay, X.T.; Goto, A. Recent development in halogen-bonding-catalyzed living radical polymerization. Polym. Chem. 2020, 11, 5559–5571. [CrossRef[

- .27 Mathur, A.M.; Moorjani, S.K.; Scranton, A.B. Methods for synthesis of hydrogel networks: A review. J. Macromol. Sci. Rev. Macromol. Chem. Phys. 1996, 36, 405–430. [CrossRef]
- .28 Szczotka-Flynn, L.B.; Imamura, Y.; Chandra, J.; Yu, C.; Mukherjee, P.K.; Pearlman, E.; Ghannoum, M.A. Increased resistance of contact lens-related bacterial biofilms to antimicrobial activity of soft contact lens care solutions. Cornea 2009, 28, 918–926. [CrossRef[
- .29 Szczotka-Flynn, L.B.; Pearlman, E.; Ghannoum, M. Microbial contamination of contact lenses, lens care solutions, and their accessories: A literature review. Eye Contact Lens 2010, 36, 116–129. [CrossRef]
- .30 Carnt, N.; Samarawickrama, C.; White, A.; Stapleton, F. The diagnosis and management of contact lens-related microbial keratitis.
- Clin. Exp. Optom. 2017, 100, 482–493. [CrossRef]
- .31 Baghbadorani, M.A.; Bigham, A.; Rafienia, M.; Salehi, H. A ternary nanocomposite fibrous scaffold composed of poly(ε- caprolactone)/Gelatin/Gehlenite (Ca2Al2SiO7): Physical, chemical, and biological properties in vitro. Polym. Adv. Technol. 2021, 32, 582–598. [CrossRef]
- .32 Bigham, A.; Aghajanian, A.H.; Movahedi, M.; Sattary, M.; Rafienia, M.; Tayebi, L. A 3D nanostructured calcium-aluminum-silicate scaffold with hierarchical meso-macroporosity for bone tissue regeneration: Fabrication, sintering behavior, surface modification and in vitro studies. J. Eur. Ceram. Soc. 2021, 41, 941–962. [CrossRef]
- .33 Foroughi, F.; Hassanzadeh-Tabrizi, S.A.; Bigham, A. In situ microemulsion synthesis of hydroxyapatite-MgFe nanocomposite as a magnetic drug delivery system. Mater. Sci. Eng. C 2016, 68. [CrossRef]
- .34 Sattary, M.; Kefayat, A.; Bigham, A.; Rafienia, M. Polycaprolactone/Gelatin/Hydroxyapatite nanocomposite scaffold seeded with Stem cells from human exfoliated deciduous teeth to enhance bone repair: In vitro and in vivo studies. Mater. Technol. 2020. [CrossRef]
- .35 Hadidi, M.; Bigham, A.; Saebnoori, E.; Hassanzadeh-Tabrizi, S.A.; Rahmati, S.; Alizadeh, Z.M.; Nasirian, V.; Rafienia, M. Electrophoretic-deposited hydroxyapatite-copper nanocomposite as an antibacterial coating for biomedical applications. Surf. Coat. Technol. 2017. [CrossRef]
- .36 Filion, T.M.; Li, X.; Mason-Savas, A.; Kreider, J.M.; Goldstein, S.A.; Ayers, D.C.; Song, J. Elastomeric Osteoconductive Synthetic Scaffolds with Acquired Osteoinductivity Expedite the Repair of Critical Femoral Defects in Rats. Tissue Eng. Part A 2010, 17, 503–511. [CrossRef]
- .37 Huang, J.; Zhao, D.; Dangaria, S.J.; Luan, X.; Diekwisch, T.G.H.; Jiang, G.; Saiz, E.; Liu, G.; Tomsia, A.P. Combinatorial design of hydrolytically degradable, bone-like biocomposites based on PHEMA and hydroxyapatite. Polymer 2013, 54, 909–919. [CrossRef[
- .38 Huang, J.; Ten, E.; Liu, G.; Finzen, M.; Yu, W.; Lee, J.S.; Saiz, E.; Tomsia, A.P. Biocomposites of pHEMA with HA/ β -TCP (60/40) for bone tissue engineering: Swelling, hydrolytic degradation, and in vitro behavior. Polymer 2013, 54, 1197–1207. [CrossRef[
- .39 Rastegar, S.; Mehdikhani, M.; Bigham, A.; Poorazizi, E.; Rafienia, M. Poly glycerol sebacate/polycaprolactone/carbon quantum dots fibrous scaffold as a multifunctional platform for cardiac tissue engineering. Mater. Chem. Phys. 2021, 266, 124543. [CrossRef[
- .40 Shams, M.; Karimi, M.; Heydari, M.; Salimi, A. Nanocomposite scaffolds composed of Apacite (apatite-calcite) nanostructures, poly (ε-caprolactone) and poly (2-hydroxyethylmethacrylate): The effect of nanostructures on physico-mechanical properties and osteogenic differentiation of human bone marrow me. Mater. Sci. Eng. C 2020, 117, 111271. [CrossRef[

- Mehrabi, T.; Mesgar, A.S.; Mohammadi, Z. Bioactive Glasses: A Promising Therapeutic Ion Release Strategy for Enhancing Wound Healing. ACS Biomater. Sci. Eng. 2020, 6, 5399–5430. [CrossRef]
- Rezvani Ghomi, E.; Khalili, S.; Nouri Khorasani, S.; Esmaeely Neisiany, R.; Ramakrishna, S. Wound dressings: Current advances and future directions. J. Appl. Polym. Sci. 2019, 136, 47738. [CrossRef[
- .43 Xu, Z.; Han, S.; Gu, Z.; Wu, J. Advances and Impact of Antioxidant Hydrogel in Chronic Wound Healing. Adv. Healthc. Mater.

].1901502 **(9 (2020CrossRef**[

- .44 Jain, A.; Tripathi, M.; Prajapati, S.K.; Raichur, A.M. Biopolymer Matrix Composite for Drug Delivery Applications in Cancer. In Reference Module in Materials Science and Materials Engineering; Elsevier: Amsterdam, The Netherlands, 2021; ISBN 978-0-12-803581-8.
- .45 Jiang, X.; He, C.; Lin, W. Supramolecular metal-based nanoparticles for drug delivery and cancer therapy. Curr. Opin. Chem. Biol.

] .153–143 **.**61 **.**2021CrossRef[

- .46 Baeza, A.; Ruiz-Molina, D.; Vallet-Regí, M. Recent advances in porous nanoparticles for drug delivery in antitumoral applications: Inorganic nanoparticles and nanoscale metal-organic frameworks. Expert Opin. Drug Deliv. 2017, 14, 783–796. [CrossRef]
- Zou, Y.; Huang, B.; Cao, L.; Deng, Y.; Su, J. Tailored Mesoporous Inorganic Biomaterials: Assembly, Functionalization, and Drug Delivery Engineering. Adv. Mater. 2021, 33, 2005215. [CrossRef] [PubMed]
- .48 Guido, C.; Maiorano, G.; Cortese, B.; D'Amone, S.; Palamà, I.E. Biomimetic Nanocarriers for Cancer Target Therapy. Bioengineering

] .111 **.**7 **.**2020CrossRef[

- .49 Moradi Kashkooli, F.; Soltani, M.; Souri, M. Controlled anti-cancer drug release through advanced nano-drug delivery systems: Static and dynamic targeting strategies. J. Control. Release 2020, 327, 316–349. [CrossRef]
- Li, H.; Liu, K.; Williams, G.R.; Wu, J.; Wu, J.; Wang, H.; Niu, S.; Zhu, L.M. Dual temperature and pH responsive nanofiber formulations prepared by electrospinning. Colloids Surf. B Biointerfaces 2018, 171, 142–149. [CrossRef] [PubMed]
- .51 Ansari, M.; Bigham, A.; Hassanzadeh Tabrizi, S.A.; Abbastabar Ahangar, H. Copper-substituted spinel Zn-Mg ferrite nanoparticles as potential heating agents for hyperthermia. J. Am. Ceram. Soc. 2018. [CrossRef]
- .52 Chouhan, R.; Bajpai, A.K. An in vitro release study of 5-fluoro-uracil (5-FU) from swellable poly-(2-hydroxyethyl methacrylate) (PHEMA) nanoparticles. J. Mater. Sci. Mater. Med. 2009, 20, 1103–1114. [CrossRef[
- .53 Chouhan, R.; Bajpai, A.K. Real time in vitro studies of doxorubicin release from PHEMA nanoparticles. J. Nanobiotechnol. 2009

].5 ·7CrossRef[

.54 Kumar, S.S.D.; Surianarayanan, M.; Vijayaraghavan, R.; Mandal, A.B.; MacFarlane, D.R. Curcumin loaded poly (2- hydroxyethyl methacrylate) nanoparticles from gelled ionic liquid—In vitro cytotoxicity and anti-cancer activity in SKOV-3 cells. Eur. J. Pharm. Sci. 2014, 51, 34–44. [CrossRef]

- Guo, J.; Hong, H.; Chen, G.; Shi, S.; Nayak, T.R.; Theuer, C.P.; Barnhart, T.E.; Cai, W.; Gong, S. Theranostic Unimolecular Micelles Based on Brush-Shaped Amphiphilic Block Copolymers for Tumor-Targeted Drug Delivery and Positron Emission Tomography Imaging. ACS Appl. Mater. Interfaces 2014, 6, 21769–21779. [CrossRef]
- .56 Polo, L.; Gómez-Cerezo, N.; García-Fernández, A.; Aznar, E.; Vivancos, J.; Arcos, D.; Vallet-Regí, M.; Martínez-Máñez, R. Mesoporous Bioactive Glasses Equipped with Stimuli-Responsive Molecular Gates for Controlled Delivery of Levofloxacin against Bacteria. Chem. Eur. J. 2018, 24, 18944–18951. [CrossRef]
- .57 Cinay, G.E.; Erkoc, P.; Alipour, M.; Hashimoto, Y.; Sasaki, Y.; Akiyoshi, K.; Kizilel, S. Nanogel-Integrated pH-Responsive Composite Hydrogels for Controlled Drug Delivery. ACS Biomater. Sci. Eng. 2017, 3, 370–380. [CrossRef] [PubMed[
- .58 Orakdogen, N.; Celik, T. Ion-stimuli responsive dimethylaminoethyl methacrylate/hydroxyethyl methacrylate copolymeric hydrogels: Mutual influence of reaction parameters on the swelling and mechanical strength. J. Polym. Res. 2016, 23, 57. [CrossRef[
- .59 Roointan, A.; Farzanfar, J.; Mohammadi-Samani, S.; Behzad-Behbahani, A.; Farjadian, F. Smart pH responsive drug delivery system based on poly(HEMA-co-DMAEMA) nanohydrogel. Int. J. Pharm. 2018, 552, 301–311. [CrossRef]
- .60 Weaver, J.V.M.; Bannister, I.; Robinson, K.L.; Bories-Azeau, X.; Armes, S.P.; Smallridge, M.; McKenna, P. Stimulus-Responsive Water-Soluble Polymers Based on 2-Hydroxyethyl Methacrylate. Macromolecules 2004, 37, 2395–2403. [CrossRef]
- .61 Ribeiro, A.; Veiga, F.; Santos, D.; Torres-Labandeira, J.J.; Concheiro, A.; Alvarez-Lorenzo, C. Bioinspired Imprinted PHEMA- Hydrogels for ocular delivery of carbonic anhydrase inhibitor drugs. Biomacromolecules 2011, 12, 701–709. [CrossRef]
- .62 Li, R.; Guan, X.; Lin, X.; Guan, P.; Zhang, X.; Rao, Z.; Du, L.; Zhao, J.J.; Rong, J.; Zhao, J.J. Poly (2-hydroxyethyl methacrylate)/ β cyclodextrin-hyaluronan contact lens with tear protein adsorption resistance and sustained drug delivery for ophthalmic diseases.

Acta Biomater. 2020, 110, 105–118. [CrossRef]

- .63 Ulu, A.; Balcioglu, S.; Birhanli, E.; Sarimeseli, A.; Keskin, R.; Koytepe, S.; Ates, B. Poly (2-hydroxyethyl methacrylate)/boric acid composite hydrogel as soft contact lens material: Thermal, optical, rheological, and enhanced antibacterial properties.
- J. Appl. Polym. Sci. 2018, 135, 46575. [CrossRef]
- Lynch, C.R.; Kondiah, P.P.D.; Choonara, Y.E.; du Toit, L.C.; Ally, N.; Pillay, V. Hydrogel Biomaterials for Application in Ocular Drug Delivery. Front. Bioeng. Biotechnol. 2020, 8. [CrossRef]
- .65 Passos, M.F.; Dias, D.R.C.; Bastos, G.N.T.; Jardini, A.L.; Benatti, A.C.B.; Dias, C.G.B.T.; Maciel Filho, R. pHEMA hydrogels.
- J. Therm. Anal. Calorim. 2016, 125, 361–368. [CrossRef]
- .66 Alipour, F.; Khaheshi, S.; Soleimanzadeh, M.; Heidarzadeh, S.; Heydarzadeh, S. Contact Lens-related Complications: A Review.
- J. Ophthalmic Vis. Res. 2017, 12, 193–204. [CrossRef]
- .67 Khan, S.A.; Lee, C.-S. Recent progress and strategies to develop antimicrobial contact lenses and lens cases for different types of microbial keratitis. Acta Biomater. 2020, 113, 101–118. [CrossRef]

- .68 Rich Kirkner, J&J's Plans for Smart & 3D Printable Contact Lenses. Available online: https://ois.net/jjs-plans-for-smart-3d-printable-contact-lenses/ (accessed on 8 June 2016.(
- .69 Rezvani Ghomi, E.; Khosravi, F.; Neisiany, R.E.; Singh, S.; Ramakrishna, S. Future of additive manufacturing in healthcare.

Curr. Opin. Biomed. Eng. 2021, 17, 100255. [CrossRef]

- .70 Rahic´, O.; Tucak, A.; Omerovic´, N.; Sirbubalo, M.; Hindija, L.; Hadžiabdic´, J.; Vranic´, E. Novel Drug Delivery Systems Fighting Glaucoma: Formulation Obstacles and Solutions. Pharmaceutics 2021, 13, 28. [CrossRef]
- .71 Ciolino, J.B.; Dohlman, C.H.; Kohane, D.S. Contact lenses for drug delivery. Semin. Ophthalmol. 2009, 24, 156–160. [CrossRef]

]PubMed[

- .72 García-Millán, E.; Koprivnik, S.; Otero-Espinar, F.J. Drug loading optimization and extended drug delivery of corticoids from pHEMA based soft contact lenses hydrogels via chemical and microstructural modifications. Int. J. Pharm. 2015, 487, 260–269. [CrossRef] [PubMed]
- .73 Andrade-Vivero, P.; Fernandez-Gabriel, E.; Alvarez-Lorenzo, C.; Concheiro, A. Improving the loading and release of NSAIDs from pHEMA hydrogels by copolymerization with functionalized monomers. J. Pharm. Sci. 2007, 96, 802–813. [CrossRef]
- .74 Li, C.-C.; Chauhan, A. Modeling Ophthalmic Drug Delivery by Soaked Contact Lenses. Ind. Eng. Chem. Res. 2006, 45, 3718–3734.

[CrossRef]

- .75 Bigham, A.; Saudi, A.; Rafienia, M.; Rahmati, S.; Bakhtiyari, H.; Salahshouri, F.; Sattary, M.; Hassanzadeh-Tabrizi, S.A. Elec- trophoretically deposited mesoporous magnesium silicate with ordered nanopores as an antibiotic-loaded coating on surface- modified titanium. Mater. Sci. Eng. C 2019, 96, 765–775. [CrossRef]
- .76 Wei, N.; Dang, H.; Huang, C.; Sheng, Y. Timolol loaded microemulsion laden silicone contact lens to manage glaucoma: In vitro and in vivo studies. J. Dispers. Sci. Technol. 2020, 42, 1–9. [CrossRef[
- .77 Xu, J.; Ge, Y.; Bu, R.; Zhang, A.; Feng, S.; Wang, J.; Gou, J.; Yin, T.; He, H.; Zhang, Y.; et al. Codelivery of latanoprost and timolol from micelles-laden contact lenses for the treatment of glaucoma. J. Control. Release 2019, 305, 18–28. [CrossRef]
- .78 Behl, G.; Iqbal, J.; O'Reilly, N.J.; McLoughlin, P.; Fitzhenry, L. Synthesis and Characterization of Poly (2-hydroxyethylmethacrylate) Contact Lenses Containing Chitosan Nanoparticles as an Ocular Delivery System for Dexamethasone Sodium Phosphate. Pharm. Res. 2016, 33, 1638–1648. [CrossRef]
- .79 Xu, J.; Li, X.; Sun, F. Cyclodextrin-containing hydrogels for contact lenses as a platform for drug incorporation and release.

Acta Biomater. 2010, 6, 486–493. [CrossRef]

- .80 Dos Santos, J.F.R.; Alvarez-Lorenzo, C.; Silva, M.; Balsa, L.; Couceiro, J.; Torres-Labandeira, J.J.; Concheiro, A. Soft contact lenses functionalized with pendant cyclodextrins for controlled drug delivery. Biomaterials 2009, 30, 1348–1355. [CrossRef]
- Ali, M.; Vaughan, A.D.; Zhang, J.; Venkatesh, S.; Byrne, M.E. Templated hydrogels for combination devices: Therapeutic contact lenses. In Proceedings of the 2009 Annual International Conference of the IEEE

- Engineering in Medicine and Biology Society, Minneapolis, MN, USA, 3–6 September 2009; Volume 2009, pp. 242–245.
- .82 Gulsen, D.; Chauhan, A. Dispersion of microemulsion drops in HEMA hydrogel: A potential ophthalmic drug delivery vehicle.
- Int. J. Pharm. 2005, 292, 95–117. [CrossRef]
- Alvarez-Lorenzo, C.; Hiratani, H.; Gomez-Amoza, J.L.; Martínez-Pacheco, R.; Souto, C.; Concheiro, A. Soft contact lenses capable of sustained delivery of timolol. J. Pharm. Sci. 2002, 91, 2182–2192. [CrossRef]
- .84 Uchida, R.; Sato, T.; Tanigawa, H.; Uno, K. Azulene incorporation and release by hydrogel containing methacrylamide propyltri- menthylammonium chloride, and its application to soft contact lens. J. Control. Release 2003, 92, 259–264. [CrossRef]
- .85 Zare, M.; Namratha, K.; Alghamdi, S.; Eissa, Y.H.; Hezam, A.; Zare, M.; Drmosh, Q.A.; Byrappa, K.; Chandrashekar, B.N.; Ramakrishna, S.; et al. Novel Green Biomimetic Approach for Synthesis of ZnO-Ag Nanocomposite; Antimicrobial Activity against Food-borne Pathogen, Biocompatibility and Solar Photocatalysis. Sci. Rep. 2019, 9, 8303–8309. [CrossRef]
- .86 Zare, M.; Namratha, K.; Byrappa, K. Green Synthesis and characterization of ZnO- Ag Nanocomposite by Thymus vulgaris.
- Int. J. Sci. Res. Sci. Technol. 2018, 4, 1636–1640.
- .87 Zare, M.; Namratha, K.; Ilyas, S.; Hezam, A.; Mathur, S.; Byrappa, K. Smart Fortified PHBV-CS Biopolymer with ZnO–Ag Nanocomposites for Enhanced Shelf Life of Food Packaging. ACS Appl. Mater. Interfaces 2019, 11, 48309–48320. [CrossRef]
- .88 Qin, G.; Zhu, Z.; Li, S.; McDermott, A.M.; Cai, C. Development of ciprofloxacin-loaded contact lenses using fluorous chemistry.

Biomaterials 2017, 124, 55–64. [CrossRef]

.89 Zare, M.; Sillanpää, M.; Ramakrishna, S. Essential role of quantum science and nanoscience in antiviral strategies for COVID-19.

Mater. Adv. 2021, 2, 2188–2199. [CrossRef]

.90 Zheng, P.; Eskandari, A.; Lu, C.; Laws, K.; Aldous, L.; Suntharalingam, K. Biophysical analysis of cancer stem cell-potent copper(ii) coordination complexes. Dalt. Trans. 2019, 48, 5892–5896. [CrossRef] [PubMed[

-

