

# Occurrence of Plasmid encoded ESBLs bla<sub>CTX-M</sub>, bla<sub>TEM</sub> genes Of E. coli isolated from Clinical cases in Maysan province

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**Abstract:** Escherichiacoli repeatedly causing urinary tract, wound and blood infection resulting in significant morbidity and mortality due to had plasmid encoded ESBLs which in turn lead to treatment failure. The present study was focused on the estimation of  $\beta$ -lactam antibiotic resistance patterns, the determination of Plasmid ESBLs represented by bla<sub>CTX-M</sub> and bla<sub>TEM</sub> gene. So a total of (291) clinical samples (urine, wound swabs, blood and seminal fluids) were included in this study. All bacterial isolates were subjected to the cultural, microscopical, and biochemical examinations methods, confirmed by API 20E and Vitek2 system. Where the results revealed that 105 of isolates were identified as E. coli. Antibiotic sensitivity was performed by using disk diffusion methods against  $\beta$ -lactam. Investigation of extended spectrum  $\beta$ -lactamase (ESBL) production for isolates was performed using Initial screening and double disc synergy method (DDST). The results showed that most isolates showed high resistance to  $\beta$ -lactam antibiotics, while all isolates were sensitive to Imipenem. The results of PCR technique were performed to detect Plasmid encoded ESBLs bla<sub>CTX-M</sub> and bla<sub>TEM</sub> genes, revealed that (100%) of E. coli isolates carried this genes for both.

**Keyword:** Escherichia coli,  $\beta$ -lactam Resistance, ESBL, bla<sub>CTX-M</sub>, bla<sub>TEM</sub> gene.

## 1. Introduction:

Plasmids are main mechanism for the spread of antibiotic resistant genes and confer traits of antibiotic resistance in bacterial populations<sup>[1]</sup>. The transfer of plasmid-mediated genes can occur either between closely related strains, or between widely related strains from diverse species or genera and can play a significant role in the mobility of resistance genes<sup>[2,3]</sup>. Resistance to  $\beta$ -lactams and other antibiotics in the Enterobacteriaceae is frequently associated with plasmid resistance determinants,  $\beta$ -Lactamase-mediated resistance is increasingly associated with plasmid-encoded extended-spectrum  $\beta$ -lactamases (ESBLs) and carbapenemases, specifically the CTX-M family of ESBLs, these enzymes are now appearing in multiple combinations of ESBLs and carbapenemases, thereby conferring resistance to virtually all  $\beta$ -lactam antibiotics<sup>[4,5]</sup>. Therefore, Plasmids have important role behind the success of the ESBL genes as they both mediate transfer and immobilize maintenance of these genes in new hosts<sup>[6]</sup>.

## 2. MATERIALS AND METHODS

### 2.1. Bacterial samples collection and identification:

A total of 291 samples, were collected from different clinical cases (urine, wound swabs, blood and seminal fluid) from main Hospitals in Maysan Province during the period from October 2018 till the end of September 2018. The samples were immediately inoculated on blood agar and MacConkey agar, then incubated for overnight at 37°C under aerobic conditions. All E. coli was isolated and identified according to their diagnostic characteristics and then compared with their being reported in MacFaddin (2000).

The isolates were confirmatively diagnosed by API 20 E system and VITEK2 system by using VITEK®2 GN kit, then stored at maintenance medium until further tests.

### 2.2. Antimicrobial susceptibility testing

Antimicrobial Susceptibility testing for  $\beta$ -lactam group was determined by the agar disk diffusion method<sup>[7]</sup>, and the results were interpreted according to CLSI guidelines<sup>[8]</sup>. The following antibiotic disks were used: Ampicillin 10  $\mu$ g, Piperacillin 100  $\mu$ g, Augmentin 20/10  $\mu$ g, Cefoxitin 30  $\mu$ g, Ceftazidime 30  $\mu$ g, Cefotaxime 30  $\mu$ g, Ceftriaxone 30  $\mu$ g, Cefipime 30  $\mu$ g, Aztreonam 30  $\mu$ g, Imipenem 10  $\mu$ g, (Bioanalyse, Turkey).

### 2.3. Phenotypic detection of ESBLs

All  $\beta$ -lactamase producing bacterial isolates were assayed for ESBL production by initial screening test according to CLSI guidelines<sup>[8]</sup>. The isolates showing resistance to one or more third generation Cephalosporins (3GCs) were tested for ESBL production by Double Disc Synergy Test (DDST)<sup>[9]</sup>.

### 2.4. Extraction of Bacterial DNA Plasmid

Plasmid extraction was done toward all E. coli isolates according to AccuPrep® Plasmid Mini extraction kit protocol (Bioneer, South Korea). The integrity of extracted Plasmid was tested using Agarose Gel Electrophoresis. The Plasmid DNA then subjected to monoplex PCR.

### 2.5. Molecular detection of CTX-M Gene Using PCR Technique

The protocol used in accordance with the manufacturer's instructions (Bioneer, South Korea). The sequences of primers and thermal cycler conditions are shown in the table (1). The amplification achieved products were separated in 1% agarose gels containing ethidium bromide 3  $\mu$ l (0.5  $\mu$ g), DNA ladder 100bp (Bioneer, Korea) was used for compare with. After electrophoresis, the gel was photographed under UV light as described by<sup>[10]</sup>.

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**Table (1)** Primers sequences and thermal cyclers conditions

Name	Primers sequences ( 5'-3')		Size bp	PCR Condition Temp/time Cycles No.	Reference
bla <sub>TEM</sub>	F R	-ACTGCGGCCAACTTACTTCTG -CGGGAGGGCTTACCATCTG	374	95C/5min 1X	[11]
				94C/30s 62C/30s 30X 72C/30s	
				72C/10min 1X	
bla <sub>CTX-M</sub>	F R	-CGCTTTGCGATGTGCAG -ACCGCGATATCGTTGGT	550	94C/5min1X	[12,13]
				94C/1min 55C/1min 34X 72C/1min	
				72C/5min 1X	

### 3. RESULTS AND DISCUSSION

#### 3.1. Isolation and identification of Bacterial Isolates

The results of this study showed that among 291 clinical samples 235 gave positive growth and 105 (44.7%) were identified as *E. coli*. As compared with other studies, our findings in line with studies reported by [14,15], where identified *E. coli* was (44%) for both, while the result of present study was contrary with study conducted by [16] where the result was (22.27%), and higher than other studies reported in India and Ethiopia implemented by [17,18], where the results were (21.96%) and (25.4%) respectively.

#### 3.2. The antibiotic susceptibility pattern

The resistance patterns of *E. coli* towards various antibiotics were determined using disc diffusion method. Data in (Table 2) exhibited that isolates of *E. coli* have the highest level of resistance to Ampicillin where up to (98.1%) were resist to this

antibiotic, followed by (96.2%) for Pipracillin and (93.3%) for Ceftazidime, Cefpodoxime, and Cefotaxime was (94.3%), whereas in (92.4%) was the resistance for Cefepime, while to Amoxicillin/Clavulanic acid the percent of resistance was (90.5%). The resistance to the Ceftriaxone was (87.6%), (86.7%) for Cefoxitin and Aztreonam (81.9%). On the other hand this study recorded that all isolates (100%) were sensitive to Imipenem antibiotic. Over all these results were directly line with local studies conducted by [19], in Iraq, also in compatible with [20], where they found high resistant to  $\beta$ -lactam antibiotic and they considered the antibiotic Imipenem and Meropenem should be preferred drugs for *E. coli* infection isolated from clinical samples. The observed high resistance rates in most antibiotic may be due to uncontrolled consumption, consequence of easy access to inefficient and cheap antibiotics moreover could be justified by insufficient adherence to guidelines for infection control as well as inappropriate use of antibiotics.

**Table(2):** Results of Antibiotics susceptibility test for *Escherichia. coli* isolated from different clinical cases (n=105).

No	Antibiotic	Sensitive n (100%)	Intermediated n (100%)	Resistance n (100%)
1	Ampicillin	2 (1.9)	-	103 (98.1)
2	Pipracillin	4 (3.8)	-	101 (96.2)
3	Amoxicillin/ Clavulanic acid	8 (7.6)	2 (1.9)	95 (90.5)
4	Cefoxitin	8 (7.6)	6 (5.7)	91 (86.7)
5	Ceftazidime	7 (6.7)	-	98 (93.3)
6	Ceftraxone	11 (10.5)	2 (1.9)	92 (87.6)
7	Cefotaxime	6 (5.7)	-	99 (94.3)
8	Cefepime	8 (7.6)	-	97 (92.4)
9	Aztronam	11 (10.5)	8 (7.6)	86 (81.9)
10	Imipenem	105 (100)	-	-

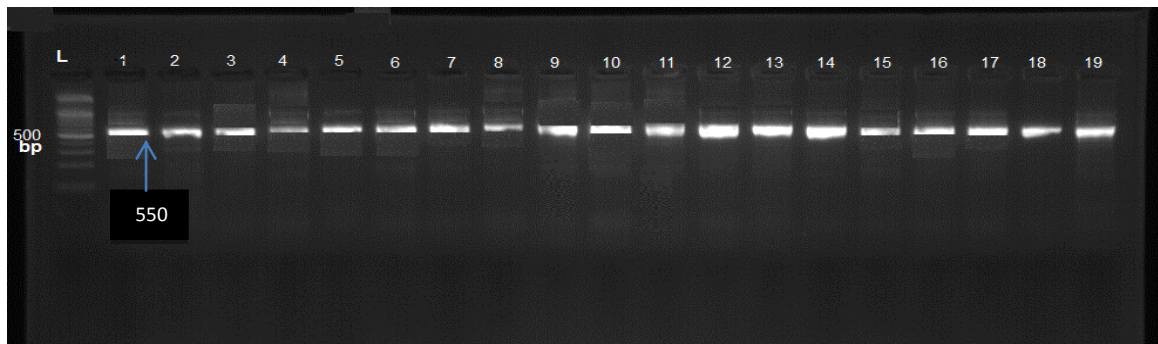
#### 3.3. Molecular detection of Plasmid encoded ESBL genes

[21] reported that plasmid carried ESBL-associated genes were prevalent in *E. coli*. Also study conducted by [22] found high prevalence of plasmid-mediated ESBLs was detected among *E. coli* in Diabetic foot infections (DFIs) in Egypt. In addition to [23] who mentioned in their studies that CTX-M  $\beta$ -

lactamases form a new and rapidly rising family of plasmid-mediated ESBLs that currently replace the transformed TEM or SHV ESBL families with high greater penetration into *E. coli*. In current study the result revealed that (100%) of *E. coli* carried bla<sub>CTX-M</sub> gene on plasmid as shown in figure (1), these findings were greatly similar to study for [22,24] where the results were (100%) for both, and in

concordance with study of<sup>[25]</sup> who found that **(86.7%0)** of *E. coli* carried **bla<sub>CTX-M</sub>** gene on plasmid. The elevated rate of CTX-M  $\beta$ -lactamases in *E. coli* isolates suggest that the horizontal transfer of **bla<sub>CTX-M</sub>** genes mediated by plasmid

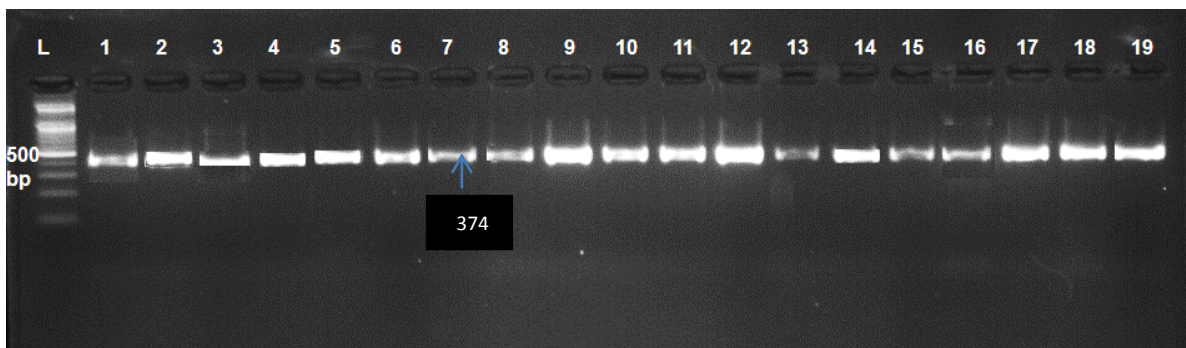
and/ or mobile genetic element, contributes to ease with which these enzymes are spreading in *E. coli* isolates and the dissemination of **CTX-M** enzymes.



**Figure (1):** Agarose gel electrophoresis of PCR plasmid encoded **bla<sub>CTX-M</sub>** gene, amplicon (550bp), where L: ladder (100bp), Lane(1-19) positive results, the gel stained by ethidium bromide (0.5  $\mu$ g/ml) and ran at (65) volts for one hour.

Current study exhibited that **(100%)** of *E. coli* isolates had **bla<sub>TEM</sub>** gene on plasmid, this rate went beyond the study conducted by<sup>[26]</sup> where the findings was **(75.6%)** and other study in a Swedish achieved by<sup>[27]</sup> where the frequency of **bla<sub>CTX-M</sub>** genes was **(63%)** among *E. coli* isolates, while these results contrary to the findings of<sup>[22,28,25,24]</sup> where the ratio reached to **(50%)**, **(56.8%)** **(32.5%)** and **(34.6%)** respectively.<sup>[29]</sup> explained that TEM type ESBLs are the first

plasmid-mediated  $\beta$ -lactamase that is often found in general of Enterobacteriaceae. Additionally,<sup>[30]</sup> who mentioned that TEM is a broad spectrum  $\beta$ -lactamase that is always combined with CTX-M in the same plasmid and the combinations of these genes are frequently seen in the ESBL producing strains, this conclusion may be one cause of prevalence **bla<sub>TEM</sub>** gene in our survey.



**Figure (2):** Agarose gel electrophoresis of PCR plasmid encoded **bla<sub>TEM</sub>** amplicon (374bp), where L: ladder (100bp), Lane(1-19) positive results, the gel stained by ethidium bromide (0.5  $\mu$ g/ml) and ran at (65) volts for one hour.

#### 4. Conclusion

In this study isolated *E. coli* showed high levels of resistance to most antibiotics of  $\beta$ -lactam group and considered as multidrug resistant bacteria. Furthermore, *E. coli* produced **bla<sub>TEM</sub>** and **bla<sub>CTX-M</sub>** genes carried on Plasmid in high rate of occurrence reached to (100%) for both. High prevalence of plasmid-mediated ESBLs was detected among clinical cases in Maysan province. Therefore, new guidelines should be undertaken in this area to restrict or prohibit the misuse and abuse of antimicrobial agents. The presence of plasmid-mediated antibiotic resistance encoded ESBLs genes shows that these genes can be disseminated to bacteria of the same or different species, and can play a significant role in the mobility of resistance genes.

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#### References

- [1]. S. S.Sandhu, "Recombinant DNA Technology". I.K. International Publishing House PVT. Ltd., New Delhi. India. pp. 59-80, 2010
- [2]. B. Ray, and A.Bhunia, "Fundamental Food Microbiology". 5th ed. CRC Press. Taylor and Francis Group, USA, pp.139-156, 2014
- [3]. M. C. Cruz, and C. T.Hedreyda, "Detection of Plasmid-Borne  $\beta$ -Lactamase Genes in Extended-Spectrum  $\beta$ -Lactamase (ESBL) and Non-ESBL-

- Producing *Escherichia coli* Clinical Isolates", *Philipp J. Sci.* vol. 146, no. 2, pp. 167-175, 2017.
- [4]. K. Bush, "Alarming  $\beta$ -lactamase-mediated resistance in multidrug-resistant Enterobacteriaceae", *Curr. Opin. Microbiol.*, vol. 13, no. 5: pp. 558-564. 2010.
- [5]. F. F. Reinthaler, G. Feierl, H. Galler, D. Haas, E. Leitner, F. Mascher, A. Melkes, J. Posch, I. Winter, G. Zarfel, and E. Marth, "ESBL-producing *Escherichia coli* in Austrian sewage sludge", *Water Res.*, vol. 2010, no. 44, pp. 1981-1985. 2010.
- [6]. A. V. L. Søråas, "Extended spectrum  $\beta$ -lactamase producing Enterobacteriaceae, Aspects on carriage, infection and treatment". MSc, thesis of Department of Medical Microbiology, University of Oslo, 2014.
- [7]. A. W. Bauer, W. M. Kirby, J. C. Sherris, and M. Turck, "Antibiotic susceptibility testing by standard single disc method", *Am J Clin Pathol.* Vol. 45, no. 4, pp. 493-496. 1966.
- [8]. CLSI, Clinical and Laboratory Standards Institute, "Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically" approved standard M100-27th ed. Wayne, PA, USA, pp. 282, 2017.
- [9]. B. Bedenić, J. Vraneš, Z. Bošnjak, T. Marijan, A., Mlinarić-Džepina, T. Kukovec, J. Knežević, M. Anušić, N. Beader, P. Barl, V. Leskovar, and S. Kalenić, "Emergence of CTX-M group 1 extended-spectrum  $\beta$ -lactamase-producing *Klebsiella pneumoniae* strains in the community", *Med. Glas.*, vol. 7, no. 1, pp. 32-39, 2010.
- [10]. A. K. Mishra, K. Sharma, R. S. Misra, "Isozyme and PCR-based genotyping of epidemic *Phytophthora colocasiae* associated with taro leaf blight", *Arch Phytopathology Plant Protect.*, vol. 43, no. 14: pp. 1367-1380, 2010.
- [11]. K. S. Kaye, H. S. Gold, M. J. Schwaber, L. Venkataraman, L., Y. Qi, P. C. De Girolami, and F. C. Tenover, "Variety of  $\beta$ -lactamases produced by amoxicillin-clavulanate-resistant *Escherichia coli* isolated in the northeastern United States", *Antimicrob. Agents. chemother.*, vol. 48, no. 5, pp. 1520-1525. 2004.
- [12]. D. L. Paterson, "Resistance in Gram-negative bacteria: Enterobacteriaceae". *Am J Med.*, vol. 119, pp. S20-S28, 2006.
- [13]. K. M. Hujer, A. M. Hujer, E. A. Hulten, S. Bajaksouzian, J. M. Adams, C. J. Donskey, and J. M. Thomson, "Analysis of antibiotic resistance genes in multidrug-resistant *Acinetobacter* sp. isolates from military and civilian patients treated at the Walter Reed Army Medical Center", *Antimicrob. agents. chemother.*, vol. 50, no. 12, pp. 4114-4123, 2006.
- [14]. N. Singh, D. Pattnaik, D. K. Neogi, J. Jena, and B. Mallick, "Prevalence of ESBL in *Escherichia coli* isolates among ICU patients in a tertiary care hospital", *J Clin Diag Res*, vol. 10, no. 9, pp. DC19-DC22, 2016.
- [15]. B. Andrew, A. Kagirita, and J. Bazira, "Prevalence of extended-spectrum  $\beta$ -lactamase-producing microorganisms in patients admitted at KRRH, Southwestern Uganda", *Int. J. Microbiol.*, vol. 2017, 2017.
- [16]. A. H. Al-Jelehaw, "Molecular Characterization of Antibacterial Resistant Genes in *Escherichia coli* Isolated from Clinical Samples". MSc, thesis, College of Medicine / University of Al-Qadisiyah. 2016.
- [17]. K. Akila, S. Sivasankari, S. Senthamarai, C. Anitha, M. Kumudavathi, and K. Muthulakshmi, "Prevalence of TEM and SHV Genes among Extended Spectrum  $\beta$ -Lactamase (ESBL) Producing *Escherichia coli* Isolates from Infected Diabetic Foot Ulcers in a Tertiary Care Hospital", *Int. J. Curr. Microbiol. App. Sci.*, vol. 6, no. 1, pp. 645-648, 2017.
- [18]. F. Gashe, E. Mulisa, M. Mekonnen, and G. Zeleke, "Antimicrobial resistance profile of different clinical isolates against third-generation cephalosporins", *J. Pharm.*, vol. 2018, 2018.
- [19]. H. H. Al-Hasnawy, R. H. Saleh, and B. H. Hadi, "Existence of ESBL genes in *Escherichia coli* and *Acinetobacter baumannii* isolated from different clinical specimens", *J Pharm Sci Res*, vol. 10 no. 5, pp. 1112-1117, 2018.
- [20]. T. Cebeci, D. Keskin, A. A. and Gököl, "Comparison of Antibiotic Sensitivity Pattern of *Escherichia coli* which Produce Extended Spectrum  $\beta$ -Lactamase Strains Isolated from Various Clinical Specimens in Intensive Care Unit", *Middle Black Sea Journal of Health Science*, vol. 5, no. 1, pp. 33-38, 2019.
- [21]. R. H. P. Dhillon, and J. Clark, "ESBLs: a clear and present danger?", *Crit Care Res Pract*, vol. 2012. pp. 11, 2012.
- [22]. N. A. Kamel, K. M. Aboshana, M. M. Abouelwafa, and W. N. El-tayeb, "Plasmid mediated extended spectrum  $\beta$ -lactamase producing strains of enterobacteriaceae isolated from diabetic foot infections in Egypt", *Arch Clin Microbiol.*, vol. 4, no. 4, 2013.
- [23]. N. S. Fam, and M. M. El-Damarawy, "CTX-M-15 extended spectrum  $\beta$ -lactamases detected from intensive care unit of an Egyptian medical research institute", *Res J Med Sci*, vol. 3, no. 1, pp. 84-91, 2008.
- [24]. B. Lohani, M. Thapa, L. Sharma, H. Adhikari, A. K. Sah, A. B. Khanal, and M. Aryal, "Predominance of CTX-M Type Extended Spectrum  $\beta$ -lactamase (ESBL) Producers Among Clinical Isolates of Enterobacteriaceae in a Tertiary Care Hospital, Kathmandu, Nepal", *Open Microbiol J*, vol. 13, no. 1, pp. 28-33, 2019.
- [25]. M. Haghghatpanah, A. S. M. Nejad, A. Mojtahedi, N. Amirmozafari, and H. Zeighami, "Detection of extended-spectrum  $\beta$ -lactamase (ESBL) and plasmid-borne blaCTX-M and blaTEM genes among clinical strains of *Escherichia coli* isolated from patients in the north of Iran", *J. Glob. Antimicrob. Resist.*, vol. 7, pp. 110-113, 2016.



- [26]. R. Azargun, M. R. Sadeghi, M. H. S. Barhaghi, H. S. Kafil, F. Yeganeh, M. A. Oskouee, and R. Ghotaslou, "The prevalence of plasmid-mediated quinolone resistance and ESBL-production in Enterobacteriaceae isolated from urinary tract infections". *Infectdrug resist*, vol. 11, pp. 1007-1014, 2018.
- [27]. H. Fang, F. Ataker, G. Hedin, and K. Dornbusch, "Molecular epidemiology of ESBLs among *Escherichia coli* isolates collected in a Swedish Hospital and its associated health care facilities from 2001 to 2006", *J Clin Microbiol*, vol. 46, pp. 707-712, 2008.
- [28]. M. N. Moghaddam, M. M. Forghanifard, and S. Moshrefi, "Prevalence and Molecular Characterization of Plasmid-mediated Extended-Spectrum  $\beta$ -Lactamase Genes (blaTEM, blaCTX and blaSHV) Among Urinary *Escherichia coli* Clinical Isolates in Mashhad, Iran". *Iran J Basic Med Sci*, vol. 15, no. 3, pp. 833-839, 2012.
- [29]. M. R. Mulvey, E. Bryce, D. A. Boyd, A. M. Ofner, S. Christianson, A. E. Simor, and S. Paton, "Ambler class A extended-spectrum beta-lactamase-producing *Escherichia coli* and *Klebsiella* spp. In Canadian hospitals", *Antimicrob Agents Chemother*, vol. 48, no. 4, pp. 1204-1209, 2004.
- [30]. R. Bonnet, "Growing group of extended-spectrum  $\beta$ -lactamases: the CTX-M enzymes" *Antimicrob Agents chemother*, vol. 48, no. 1, pp. 1-14, 2004.